

## Thorectandrols A and B, New Cytotoxic Sesterterpenes from the Marine Sponge *Thorectandra* Species

Romila D. Charan, Tawnya C. McKee, and Michael R. Boyd\*

Laboratory of Drug Discovery Research and Development, Center for Cancer Research, National Cancer Institute, Frederick, Maryland, 21702-1201

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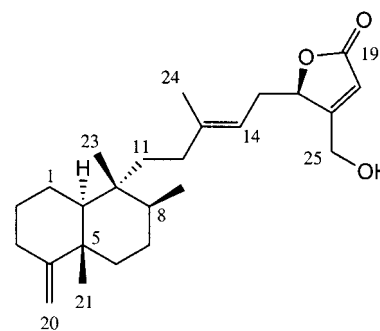
Two new sesterterpenes, thorectandrol A (**1**) and B (**2**), were isolated from extracts of the marine sponge *Thorectandra* sp. The structures were determined by extensive NMR spectral data analysis. NOE correlations were used to define the relative stereochemistry of **1** and **2**, while CD data were used to suggest their absolute stereochemistry. Both compounds **1** and **2** inhibited the growth of MALME-3M (melanoma) and MCF-7 (breast) cancer cell lines in the range 30–40  $\mu\text{g/mL}$ . The known compound palauolol (**3**) was isolated as well and was also cytotoxic.

Sesterterpene metabolites are widespread in the sponge family Thorectidae;<sup>1</sup> however, to date only five investigations on the genus *Thorectandra* have appeared in the chemical literature.<sup>2–6</sup> In the course of our continuing studies on cytotoxic constituents from marine organisms we investigated a sponge of the genus *Thorectandra* collected in Palau.

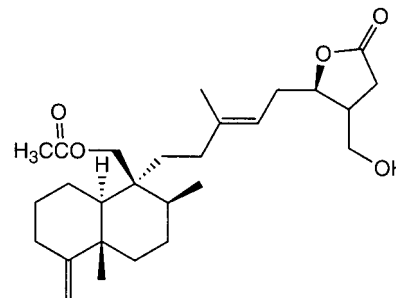
The crude organic extract of the frozen sponge showed antiproliferative activity in the melanoma and breast cancer cell lines and therefore was subjected to bioassay-guided purification. Solvent–solvent partitioning provided a cytotoxic methyl *tert*-butyl ether fraction, which was purified by reversed-phase HPLC to give thorectandrol A (**1**, 9.5 mg), thorectandrol B (**2**, 1.5 mg), and the known compound palauolol (**3**, 34.0 mg).

Thorectandrol A (**1**) was isolated as an optically active, light yellow oil. The molecular formula  $\text{C}_{25}\text{H}_{38}\text{O}_3$  was established by HRFABMS (obsd  $m/z$  387.2883 for  $[\text{M} + \text{H}]^+$ , calcd  $m/z$  387.2899), indicating seven degrees of unsaturation. The  $^{13}\text{C}$  NMR spectrum of **1** contained resonances for all 25 carbons, while a DEPT experiment revealed the presence of four methyls, 10 methylenes, five methines, and six quaternary carbons. An IR absorption band at  $1760\text{ cm}^{-1}$  indicated the likely presence of a lactone carbonyl functionality, which was supported by the presence of an ester carbonyl resonance at  $\delta$  175.4 in the  $^{13}\text{C}$  NMR spectrum. The carbonyl resonance and six additional carbon resonances between  $\delta$  103.2 and 175.9, which were assigned to three olefin groups based on COSY and HMBC data, accounted for four degrees of unsaturation and suggested the presence of three rings in **1**. A broad infrared absorption band at  $3250\text{ cm}^{-1}$  suggested the presence of an OH group.

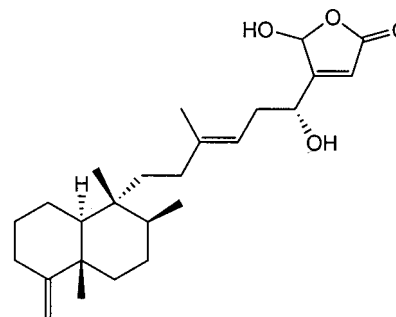
The presence of a bicyclic ring containing an exocyclic double bond and an  $\alpha,\beta$ -unsaturated butenolide moiety in **1** was apparent from detailed analysis of  $^1\text{H}$  and  $^{13}\text{C}$  NMR data together with COSY, HSQC, and HMBC spectra (Table 1). The  $^1\text{H}$  and  $^{13}\text{C}$  NMR data for the C-1 to C-14 fragment in **1** closely resembled the corresponding fragment in palauolol (**3**).<sup>7</sup> A standard series of 2D NMR experiments confirmed the assignment of C-1 to C-14. COSY correlations from H-14 ( $\delta$  4.99) to H-15 ( $\delta$  2.36 and 2.66) together with HMBC correlations from H-15 to C-14 indicated the attachment of a methylene group at C-14.



Thorectandrol A (**1**)



Thorectandrol B (**2**)



Palauolol (**3**)

H-15 in turn showed COSY correlations to H-16 ( $\delta$  5.13), and their attachment was supported by HMBC correlations in both directions. The  $^1\text{H}$  NMR spectrum of **1** displayed a one-proton resonance at  $\delta$  5.97 (H-18), which had a long-

\* To whom correspondence should be addressed. Tel: 301 846-5391. Fax: 301 846-6919. E-mail: boyd@dtpx2.ncifcrf.gov.

**Table 1.**  $^1\text{H}$  (500 MHz) and  $^{13}\text{C}$  (125 MHz) NMR Spectral Data for Thorectandrol A (**1**) and B (**2**) (MeOH- $d_4$ )

C no.	<b>1</b>			<b>2</b>	
	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (mult., $J = \text{Hz}$ )	HMBC	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (mult., $J = \text{Hz}$ )
1	22.8	1.48 (m) 1.55 (m)	C-2, C-10	23.9	1.58 (m) 1.66 (m)
2	29.8	1.23 (m) 1.89 (m)	C-3	30.2	1.28 (m) 1.90 (m)
3	34.1	2.08 (m) 2.30 (m)	C-2, C-4	34.3	2.10 (m) 2.29 (m)
4	161.7			161.7	
5	41.2			40.9	
6	38.6	1.52 (m) 1.56 (m)	C-5, C-7, C-8, C-21 C-5, C-7, C-8	31.6	1.57 (m, 2H)
7	28.6	1.43 (m, 2H)	C-5, C-6	28.5	1.55 (m, 2H)
8	37.9	1.44 (m)	C-9, C-11, C-22	37.9	1.55 (m)
9	40.3			43.8	
10	50.0	1.09 (dd, 12.2, 2.8)	C-1, C-5, C-21, C-23	50.3	1.23 (dd, 12.4, 2.7)
11	38.2	1.27 (m) 1.39 (m)	C-9, C-10, C-12 C-9, C-12	38.7	1.58 (m) 1.64 (m)
12	34.2	1.69 (m) 1.83 (m)	C-11, C-13, C-14, C-24 C-11, C-13, C-14, C-24	33.7	1.73 (m) 1.85 (m)
13	142.4			142.2	
14	116.8	4.99 (t, 7.0)	C-24	117.1	5.02 (t, 7.0)
15	31.6	2.36 (m) 2.66 (m)	C-13, C-14, C-16 C-13, C-16, C-17	31.5	2.37 (m) 2.70 (m)
16	83.1	5.13 (t, 5.0)	C-14, C-15, C-17, C-18	83.8	5.16 (t, 4.4)
17	175.9			175.8	
18	115.7	5.97 (d, 1.2)	C-19	115.7	5.97 (d, 1.8)
19	175.4			175.3	
20	103.2	4.48 (s) 4.49 (s)	C-3, C-5 C-3, C-5	103.8	4.49 (s) 4.53 (s)
21	21.4	1.05 (s, 3H)	C-4, C-5, C-6, C-10	21.4	1.05 (s, 3H)
22	16.4	0.80 (d, 6.4, 3H)	C-7, C-8, C-9	17.4	0.89 (d, 6.1, 3H)
23	18.6	0.73 (s, 3H)	C-9, C-10, C-11	67.7	4.16 (s, 2H)
24	16.6	1.62 (s, 3H)	C-12, C-13, C-14	16.7	1.65 (s, 3H)
25	59.1	4.39 (dd, 17.7, 1.3, 2H)	C-17, C-18	59.1	4.40 (dd, 17.7, 1.8, 2H)
26				173.2	
27				21.0	2.03 (s, 3H)

range COSY coupling to H-16. In addition HMBC correlations from H-16 to C-18 ( $\delta$  115.7) confirmed the assignment of C-18. The carbonyl resonance at C-19 ( $\delta$  175.4) was assigned based on a HMBC correlation from H-18 to C-19. A two proton resonance at  $\delta$  4.39 was consistent with the presence of an oxymethylene group substituted at C-17, and this was supported by HMBC correlations from H-25 to C-17. H-25 also showed long-range COSY correlations to H-18, supporting the presence of an  $\alpha,\beta$ -unsaturated butenolide moiety with an oxymethylene group substituted at C-17. Although  $\alpha,\beta$ -unsaturated butenolide moieties have been reported in many natural products,<sup>2,7-13</sup> to the best of our knowledge, only compounds isolated from *Luffariella variabilis*<sup>12</sup> and *L. geometrica*<sup>13</sup> have an oxymethylene group substituted at the  $\beta$ -carbon as found in **1**. The *E* configuration<sup>14</sup> of the  $\Delta^{13}$  double bond was inferred from the upfield  $^{13}\text{C}$  resonance at 16.6 ppm assigned to C-24 and confirmed by NOE enhancements of both H-15 protons on irradiation of H-24. One-dimensional gNOESY<sup>15</sup> experiments allowed the assignment of the relative stereochemistry of the bicyclic ring in **1**. Our assignment of the relative stereochemistry of the bicyclic ring was consistent with that proposed in the literature based on  $^{13}\text{C}$  chemical shifts.<sup>16</sup>

A molecular formula of  $\text{C}_{27}\text{H}_{40}\text{O}_5$  was established by HRFABMS for thorectandrol B (**2**). Inspection of the  $^1\text{H}$  NMR spectrum (Table 1) revealed that **2** was closely related to thorectandrol A (**1**) but contained resonances for a new oxymethylene at 4.16 ppm instead of the methyl at 0.73 ppm. In addition a new methyl singlet at 2.03 ppm indicated the presence of an acetate functionality. The  $^{13}\text{C}$  spectrum of **2** contained an extra resonance at  $\delta$  67.7, which was assigned to the oxymethylene group. The ester carbonyl of the acetate group appeared at 173.2 ppm (C-26),

and HMBC correlations from H-23 ( $\delta$  4.16) to C-26 indicated the attachment of the acetate group at C-23. Further, HMBC correlations from H-23 to C-8 and C-9 allowed placement of the oxymethylene group bearing the acetate functionality at C-9. The NMR data of the C-11 to C-19 region of thorectandrol B (**2**) were nearly identical to that for thorectandrol A (**1**) (Table 1). The *E* configuration of the  $\Delta^{13}$  double bond and the relative stereochemistry of the bicyclic ring were assigned using NOE correlations in the same manner as was done for thorectandrol A (**1**) and are as shown in **2**.

In an effort to assign the absolute stereochemistry at C-16, the CD data of thorectandrol A (**1**) and B (**2**) were compared with those of luffarin K and luffarin L.<sup>13</sup> Both thorectandrol A (**1**) and B (**2**) displayed a negative Cotton effect at  $\lambda_{\text{max}}$  212 and 210 nm, respectively, a result similar to that reported for luffarin K and L,<sup>13</sup> and hence an *R* stereochemistry was suggested for C-16 in **1** and **2**.

The absolute stereochemistry at C-16 in palauolol (**3**) has also been reported to be *R*.<sup>7</sup> The absolute stereochemistry of the bicyclic ring system of thorectandrol A (**1**) and B (**2**) was tentatively assigned by comparison of their CD spectra with CD data reported for palauolol<sup>7</sup> and data obtained by our own measurements. Thorectandrol A (**1**) and B (**2**) both showed a positive Cotton effect at  $\lambda_{\text{max}}$  197 nm, exactly the same as palauolol, and hence this supported the assignment of 5*S*, 8*S*, 9*R*, 10*S* stereochemistry in both **1** and **2**.

The major compound isolated was identified as palauolol (**3**) by comparison of its NMR data with that of published data.<sup>7</sup>

All three compounds were tested for in vitro cytotoxicity against two human tumor cell lines, MALME-3M (melanoma) and MCF-7 (breast).<sup>17</sup> Thorectandrol A (**1**) and B

(2) inhibited growth of the MALME-3M cancer cell line (IC<sub>50</sub> 40.0 and IC<sub>50</sub> 30.0 μg/mL, respectively) and the MCF-7 cancer cell line (IC<sub>50</sub> 40.0 and IC<sub>50</sub> 30.0 μg/mL, respectively). Palauolol (3) showed similar activity in both the cell lines as well (IC<sub>50</sub> 0.46 and IC<sub>50</sub> 14.2 μg/mL, respectively).

### Experimental Section

**General Experimental Procedures.** Optical rotations were recorded on a Perkin-Elmer 241 polarimeter in MeOH. UV spectra were obtained on a Beckman DU-64 spectrophotometer and FTIR spectra on a Perkin-Elmer 267 spectrophotometer. All 1D and 2D NMR spectra were recorded on a Varian Unity Inova spectrometer at 500 and 125 MHz for <sup>1</sup>H and <sup>13</sup>C, respectively, and referenced to the residual solvent signal. The number of attached protons for <sup>13</sup>C resonances were determined from a DEPT experiment. Proton-detected heteronuclear correlations were measured using HSQC (optimized for <sup>1</sup>J<sub>C-H</sub> = 140 Hz) and HMBC (optimized for <sup>n</sup>J<sub>C-H</sub> = 8.3 Hz) pulse sequences. The gNOESY experiments were carried out using a mixing time of 300 ms. Mass spectra were recorded on a JEOL SX102 spectrometer.

**Sponge Material.** The sponge was collected in 1991 from Palau by P. Colin of the Coral Reef Research Foundation, under contract for the National Cancer Institute. The sponge was collected at a depth of 10 m and is 5–15 cm in diameter. It is dark rusty red both inside and outside, and the surface is convoluted and tears easily. A voucher specimen (QCDN5079) from this collection is maintained at the Smithsonian Institution. A photograph of the sponge is provided in the Supporting Information.

**Extraction and Purification.** The frozen sponge material was ground to a coarse powder (327 g) and sequentially extracted with water followed by MeOH/CH<sub>2</sub>Cl<sub>2</sub> (1:1). The organic extract was evaporated in vacuo and dried to give a maroon solid (4.89 g). A 1.08 g portion of the cytotoxic crude extract was subjected to solvent–solvent partitioning. A portion (146 mg) of the methyl *tert*-butyl ether fraction was purified by reversed-phase HPLC (1.0 × 25 cm; C-18; 60 Å; Dynamax) using 90% (by volume) of acetonitrile in water, yielding thorectandrol A (1, 9.5 mg). Two earlier eluting sesterterpene fractions were further purified by RPHPLC using 80% and 90% (by volume) of acetonitrile in water, giving thorectandrol B (2, 1.5 mg) and palauolol (3, 34 mg), respectively. Elution of the individual components was monitored using a photodiode array detector (Waters 990).

**Cytotoxicity Assays.** Cytotoxicity assays using MALME-3M (melanoma) and MCF-7 (breast) cancer cell lines were performed as described before.<sup>17</sup>

**Thorectandrol A (1):** yellow oil; 9.5 mg, 6.51% extract wt, [α<sub>D</sub>] −15.0° (c 0.15, MeOH); CD (MeOH) λ<sub>max</sub> (log Δε) 212

(−4.1), 197 (+4.3) nm; UV (MeOH) λ<sub>max</sub> (log ε) 209 (3.94) nm; IR (film) ν<sub>max</sub> 3250 (OH), 1760 (C=O), 1630 (C=C) cm<sup>−1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Table 1; HRFABMS *m/z* obsd 387.2883 for [M + H]<sup>+</sup>, calcd for C<sub>25</sub>H<sub>39</sub>O<sub>3</sub> 387.2899.

**Thorectandrol B (2):** yellow oil; 1.5 mg, 1.03% extract wt, [α<sub>D</sub>] −19.4° (c 0.07, MeOH); CD (MeOH) λ<sub>max</sub> (log Δε) 210 (−4.5), 197 (+4.7) nm; UV (MeOH) λ<sub>max</sub> (log ε) 210 (4.05) nm; IR (film) ν<sub>max</sub> 3370 (OH), 1760 (C=O), 1630 (C=C) cm<sup>−1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR data, see Table 1; HRFABMS *m/z* obsd 445.2938 for [M + H]<sup>+</sup>, calcd for C<sub>27</sub>H<sub>41</sub>O<sub>5</sub> 445.2922.

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**Supporting Information Available:** A photograph of the sponge *Thorectandra* sp. is available free of charge via the Internet at <http://pubs.acs.org>.

### References and Notes

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